

Abstract

Monitoring of suspension cell cultures often relies on sampling followed by a staining procedure. Estimations of the cell count and cell viability are traditionally performed once a day using Trypan-Blue cell exclusion as a method of choice. This process involves manual operations and weekend work is regularly needed.

Quantitative phase imaging (QPI) is a new quantitative imaging technique that allows cell counting as well as cell viability monitoring in a continuous, label-free set-up. No need for sampling (thus eliminating the risk of contamination and the generation of toxic wastes), staining and waiting for the results generated by an off-line counter: results are available in nearly real-time over the whole run.

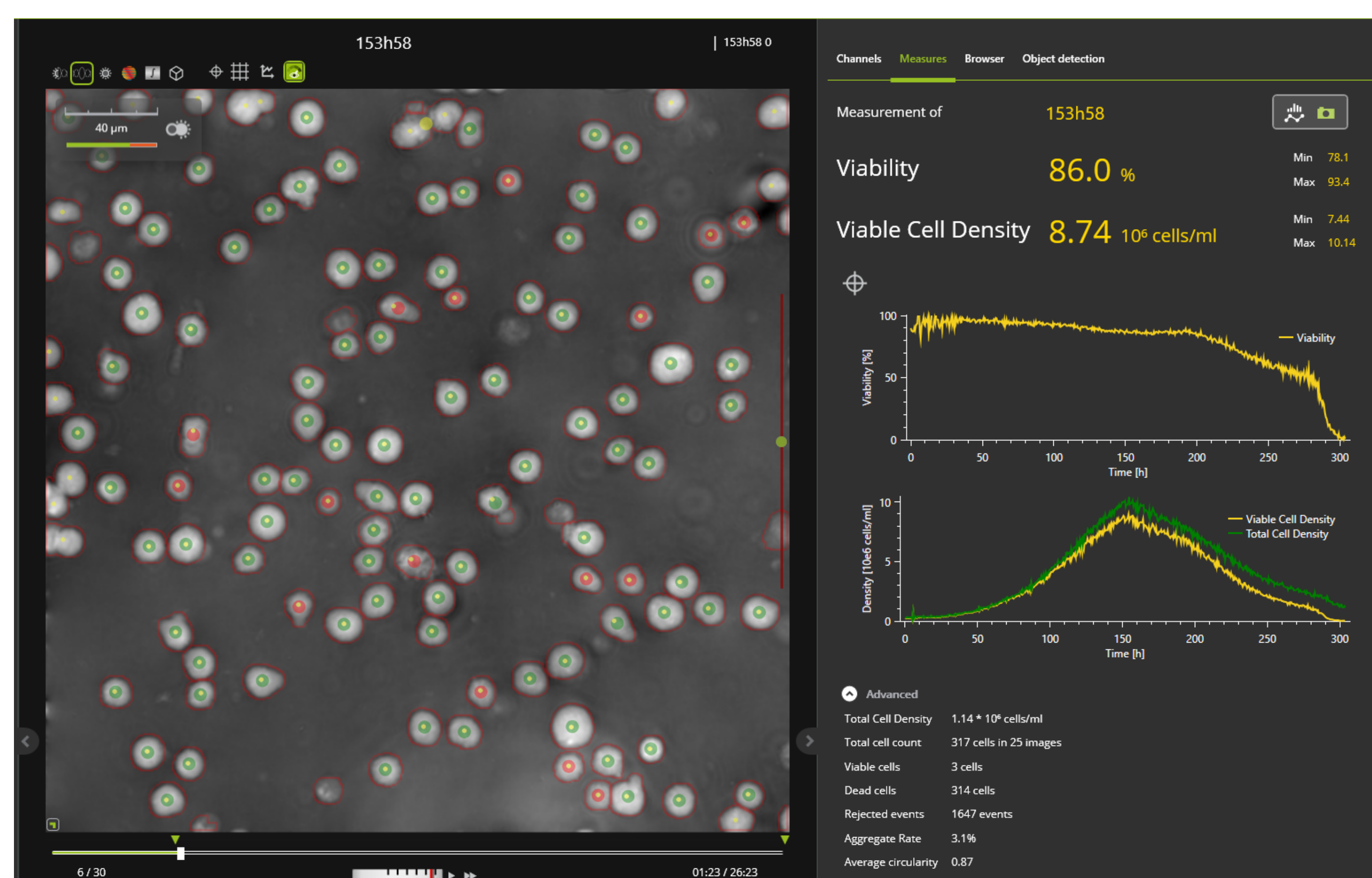
Compared to classical light microscopy, QPI offers:

- The ability to refocus images post acquisition
- The collection of quantitative phase information (optical density), covering the shape and density of an object. This quantitative phase parameter (not captured by the human eye) is the key advantage in numerous applications developed at OVIZIO.

QPI helps the operator to track the total cell density and the cell viability at any time, while the OsOne software plots the cell growth curve, live on the screen. Moreover, OsOne also shows real-time images of the cells, offering the experienced operator a convenient tool to look at the culture, live.

Results generated by an iLine F were compared with a reference methods: the Vi-Cell XR off-line counter (Beckman Coulter), automated Trypan-Blue staining. A correlation factor R^2 of 0.993 was obtained on the Viable Cell Density demonstrating that the results obtained with QPI are in line with current reference methods.

Results



Future Developments

- The availability of full data, per cell, for the whole experiment allows to **envison the use of iLine F for a PAT approach**. Indeed the large amount of data produced can be used to perform various statistical analysis on the cell population in order to define and control critical parameters of the process.
- The **iLine F can be linked with the bioreactor controller** in order to control for instance the feeding, based on the cell density, or the harvest time after a viral infection, based on the viability of the culture. The next version of the device will introduce multiplexing: OsOne will act as a **central control server** that collects data from several iLine-F simultaneously. Thus a centralized computer could perform all the calculations while only the microscope occupies valuable space in the lab.
- Furthermore, smartphones could also connect to the central control server so cell data are accessible **anywhere, anytime**.
- Finally, an API (**Application Programming Interface**) is currently in development in order to integrate into other Processes Information Management Systems.

Advantages

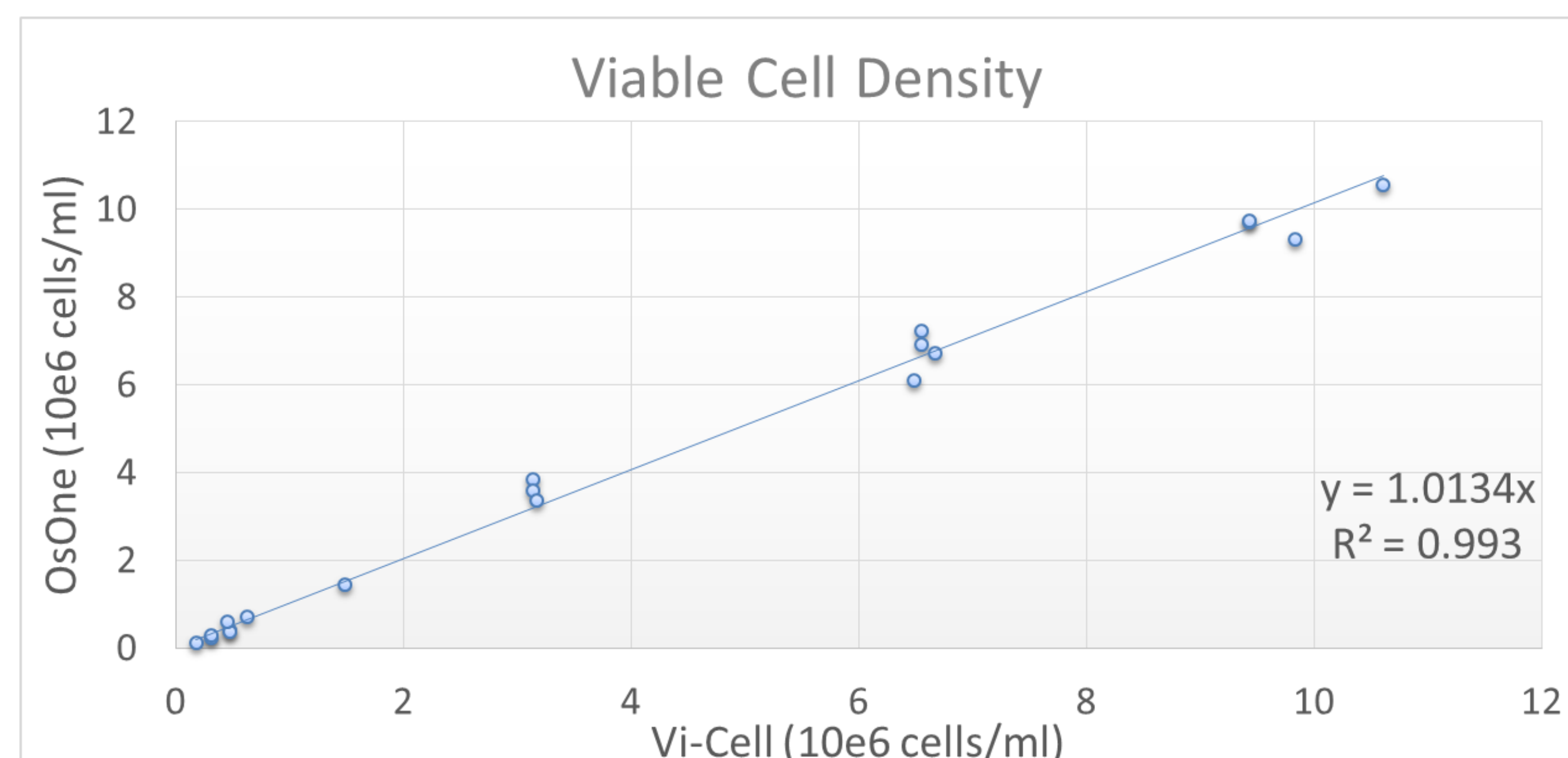
- Full traceability at single cell level
- Continuous monitoring
- User independent, improving reproducibility
- No sampling, no staining
- No toxic waste
- Ready for automation
- Increased insight in your culture process
- Adds cluster information as clusters are not disrupted during sample preparation
- More than just cell density and viability: up to 59 parameters are recorded

Materials & Methods

- iLine F microscope.
- **BioConnect:** autoclavable and disposable **closed-loop** interface with the bioreactor.
- Reusable pump engine with automated flushing (in case air bubbles or large aggregates are detected, patent pending).
- **OsOne Acquisition & Analysis software** (developed in-house by OVIZIO), version 5.9.
- Applikon 3 liters glass bioreactor controlled by ez-Control.
- CHO cells inoculated at 0.3×10^6 viable cells/mL in CD-OptiCHO™ medium (Life Technologies), batch culture for 12 days.



Conclusions

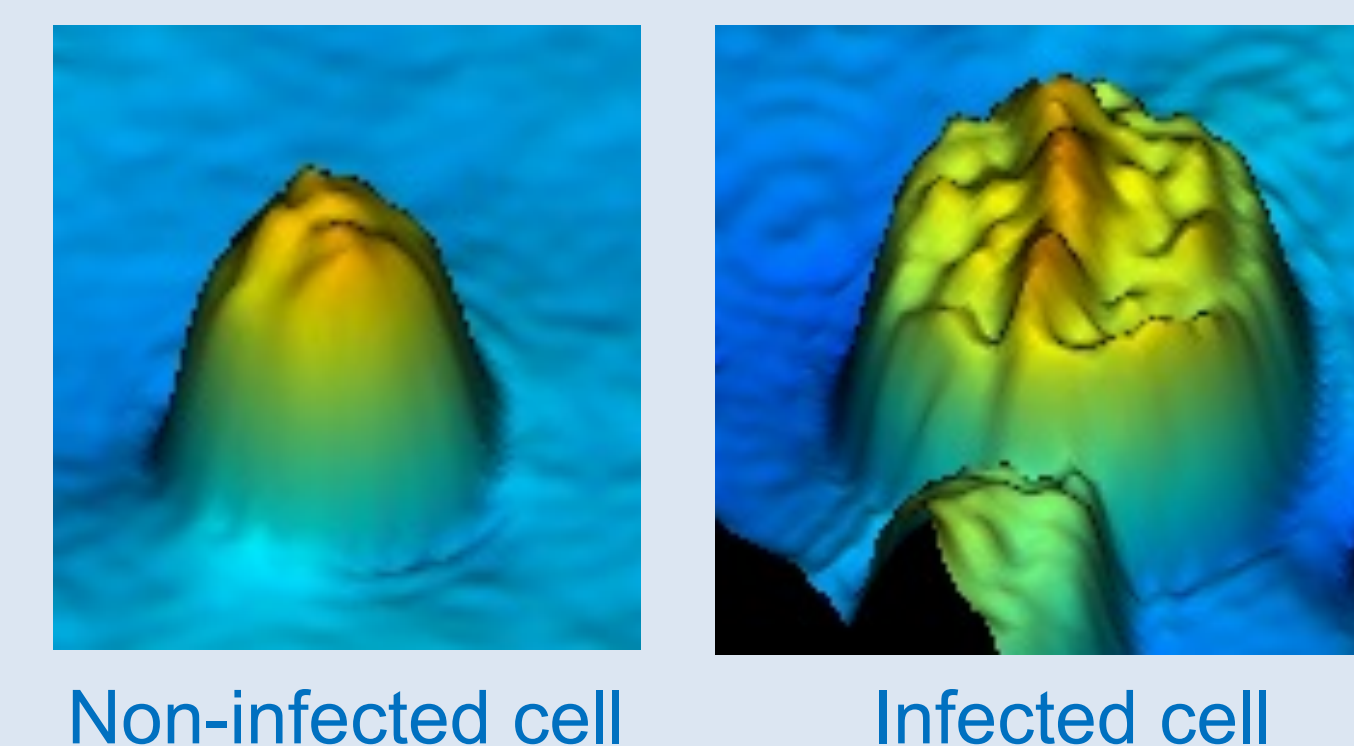


- **Very good correlation ($R^2=0.993$) of the viable cell density** compared to classic staining methods applying sampling and Trypan-Blue staining. This also proves that **the label-free approach is equivalent** to extensively validated reference methods.
- Full data per cell is available, for the whole experiment, for validation, QC, audit, etc.

Other Applications of QPI

QPI has the capability to generate an optical signature for any type of cells. As a consequence, one could envision many other applications such as:

Counting of infected cells: when infected by a virus, cells are showing a clear optical signature that can be captured by QPI. With a limited training set, a false positive rate of 8,2% and false negative rate of 7,3% were obtained.



Identification and count of blood cells: each blood cell have a particular optical signature linked to its shape, size and content.

